



## CONFERENCE 2017, Dublin, Ireland

### Applied Sheep and Goat Genetics

**Organised by a standing committee**      yes

**Date and meeting time:** 20<sup>th</sup> July 2017, 14:30 – 16:10

**Chair, name and contact email:** Gesine Lühke, Gesine.Luehken@agrar.uni-giessen.de

#### **Agenda / programme:**

<b>Time</b>	<b>Speaker</b>	<b>Title</b>
14:30	L. Drouilhet	Introgression of wool-shedding genes into the Romane breed sheep
14:45	A. Haile	Community-based sheep breeding programs in Ethiopia resulted in substantial genetic gains
15:00	L. Chantepie	Identification of two major genes affecting prolificacy in the French Noire du Velay sheep
15:15	M.R. Mousel	Genomic Regions Associated with Entropion in Columbia, Polypay, and Rambouillet Breeds of Sheep
15:30	L. Jiang	Genome-wide scan reveals NF1 locus is associated with fat tail phenotype rather than high-altitude adaptation in Asian sheep
15:45	K.M. Davenport	Investigating genetic associations with meiotic recombination in rams
16:00		Decisions on sheep and goat comparison tests for 2018/19
~16.05		Meeting ends

**Number of participants at meeting:** about 50

#### **Summary of the meeting:**

All presentations were given as scheduled. The different topics were nicely presented and interesting points were discussed by the auditorium.

The following decisions were made (no votes against):

- 1) In 2018/19, STR comparison tests for sheep and goats will be conducted.

2) Duty labs for 2018/19 will be:

Sheep CT: Labogena, France (Alexandre Vasilescu; alexandre.vasilescu@labogena.fr)

Goat CT: Laboratorio de Genética Molecular Aplicada, Animal Breeding Consulting, S.L., Spain (Amparo Martinez; amparomartinezuco@gmail.com)

3) Decisions about implementation of comparison tests for parentage testing using SNPs for sheep and/or goat will be adjourned until the next conference (Lleida, 2019).

In order to gain a picture of the current opinions about the plan to ship in future comparison test samples to participants by a central lab in Europe, the workshop attendants were asked to vote in favour or against. Result: 0 in favour, 6 against, 2 undecided (large majority did not give a vote).

### **Committee members** (the old and new committee)

<b>Chair</b>	<b>term of service</b>	<b>E mail address</b>
Gesine Lühken	2014-2019	Gesine.Luehken@agrar.uni-giessen.de

<b>Other members</b>	<b>term of service</b>	<b>E mail address</b>
Bengi Cinar Kul	2014-2019	bkul@ankara.edu.tr
Rosina Fossati	2014-2019	fossati@genia.com.uy
Remy Faugeras*	2017-2019	remy.faugeras@labogena.fr
Meng-Hua Li	2016-2019	menghua.li@ioz.ac.cn
Amparo Martinez	2014-2019	amparomartinezuco@gmail.com
Mohammad Hossein Moradi	2014-2019	hoseinmoradi@ut.ac.ir
Clementine Rodellar	2014-2019	rodellar@unizar.es
Xiaolong Wang	2014-2019	xiaolongwang@nwsuaf.edu.cn
Stephen White	2014-2019	swhite@vetmed.wsu.edu

\*substitutes Julie Ogereau since February 2017

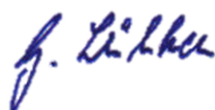
### **List of recommended markers with primer information**

- see list on ISAG homepage

### **Duty laboratory for the next comparison test**

- see decision on point 2) above

### **SIGNATURE**



**Chair**



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### ISAG ovine microsatellite (STR) panel

Marker	Forward primer sequence (5'-3')	Reverse primer sequence (5'-3')	Fragment sizes in bp (samples CT 2015-16)
AMEL	CAGCCAAACCTCCCTCTGC	CCCGCTTGGTCTTGTCTGTTGC	(X and Y)
CSRD247	GGACTTGCCAGAACTCTGCAAT	CACTGTGGTTTGTATTAGTCAGG	209-255
ETH152/D5S2	TACTCGTAGGGCAGGCTGCCTG	GAGACCTCAGGGTTGGTGATCAG	186-200
INRA005	TTCAGGCATACCCTACACCACATG	AAATATTAGCCAACTGAAAACCTGGG	125-147
INRA006	AGGAATATCTGTATCAACCGCAGTC	CTGAGCTGGGGTGGGAGCTATAAATA	110-132
INRA023	GAGTAGAGCTACAAGATAAACTTC	TAACTACAGGGTGTTAGATGAACTC	194-216
INRA063	GACCACAAAGGGATTTGCACAAGC	AAACCACAGAAATGCTTGGAAG	169-201
INRA172	CCAGGGCAGTAAAATGCATAACTG	GGCCTTGCTAGCCTCTGCAAAC	126-160
MAF065	AAAGGCCAGAGTATGCAATTAGGAG	CCACTCCTCCTGAGAATATAACATG	125-137
MAF214	AATGCAGGAGATCTGAGGCAGGGACG	GGGTGATCTTAGGGAGGTTTTGGAGG	189-265
MCM042	CATCTTTCAAAGAAGACTCCGAAAGTG	CTTGGAATCCTTCCTAACTTTCGG	87-107
MCM527	GTCCATTGCCTCAAATCAATTC	AAACCACTTGACTACTCCCCAA	164-170
OARFCB20	GGAAAACCCCATATATACCTATAC	AAATGTGTTTAAGATTCCATACATGTG	87-113



## CONFERENCE 2017, Dublin, Ireland

### ISAG caprine microsatellite (STR) panel

Marker	Forward primer sequence (5'-3')	Reverse primer sequence (5'-3')	Fragment sizes in bp (samples CT 2015-16)
<b>CSRD247</b>	GGACTTGCCAGAACTCTGCAAT	CACTGTGGTTTGTATTAGTCAGG	216-240
<b>ILSTS008</b>	GAATCATGGATTTTCTGGGG	TAGCAGTGAGTGAGGTTGGC	174-182
<b>ILSTS19</b>	AGGGACCTCATGTAGAAGC	ACTTTTGGACCCTGTAGTGC	146-152
<b>ILSTS87</b>	AGCAGACATGATGACTCAGC	CTGCCTCTTTTCTTGAGAGC	133-151
<b>INRA005</b>	TTCAGGCATACCCTACACCACATG	AAATATTAGCCAAGTGAAGTGGG	115-121
<b>INRA006</b>	AGGAATATCTGTATCAACCGCAGTC	CTGAGCTGGGGTGGGAGCTATAAATA	107-123
<b>INRA023</b>	GAGTAGAGCTACAAGATAAACTTC	TAACTACAGGGTGTGTTAGATGAACTC	195-215
<b>INRA063</b>	GACCACAAAGGGATTTGCACAAGC	AAACCACAGAAATGCTTGGAAG	171-177
<b>MAF65</b>	AAAGGCCAGAGTATGCAATTAGGAG	CCACTCCTCCTGAGAATATAACATG	117-135
<b>MCM527</b>	GTCCATTGCCTCAAATCAATTC	AAACCACTTGACTACTCCCAA	152-164
<b>OARFCB20</b>	GGAAAACCCCATATATACCTATAC	AAATGTGTTTAAGATTCCATACATGTG	95-105
<b>SRCRSP5</b>	GGACTCTACCAACTGAGCTACAAG	TGAAATGAAGCTAAAGCAATGC	163-179
<b>SRCRSP8</b>	TGCGGTCTGGTTCTGATTTAC	CCTGCATGAGAAAGTCGATGCTTAG	220-240
<b>SRCRSP23</b>	TGAACGGGTAAAGATGTG	TGTTTTTAATGGCTGAGTAG	77-103